# NEW TRITERPENES FROM THE LEAVES OF *OLEA EUROPAEA*

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Abstract—The terpene fraction from the leaves of *Olea europaea* has been shown to contain the known oleanolic and maslinic acids, together with oleanane hydroxy acids which have never been found before in nature.

### INTRODUCTION

VERY recently, we reported¹ that oleanolic acid (1a) is practically the only detectable triterpene of olive husks. Maslinic acid (1b), earlier reported² as a second triterpene of the husks, was in fact proved to be an artefact arising from oleanolic acid itself, possibly via microbiological hydroxylation occurring during the ageing of the husks. Since maslinic acid has also been isolated from other natural sources³ where its formation from oleanolic acid seemed unlikely, we extended our chemical study to the leaves of *Olea europaea* in order to check the presence of maslinic acid in other organs of the plant.

#### RESULTS

The leaves, collected in the summer from different plants, were dried at room temperature\* and then exhaustively extracted with both cold hexane and cold acetone. Hexane afforded a large fraction (5%) consisting of fats which was not further considered. The acetone extract (2%) was instead submitted to several chromatographic fractionations which gave more fats and waxes together with two terpene fractions mainly consisting of oleanolic acid (1a) and of maslinic acid (1b) respectively.

The crude maslinic acid (1b), purified through chromatography and crystallization from MeOH, gave the pure product m.p.  $267-70^{\circ}$ ,  $[\alpha]_D + 40^{\circ}$ , identical with authentic material. Evaporation of the mother liquors of crystallization of 1b, followed by treatment of the resulting solid with ethereal diazomethane, afforded an oil whose NMR examination evidenced the marked enrichment of a second substance. Crystallization of the mixture from hexane afforded a solid m.p.  $183-7^{\circ}$ ,  $[\alpha]_D - 28^{\circ}$  C<sub>31</sub>H<sub>50</sub>O<sub>4</sub>, isomeric with methyl maslinate (1c). The NMR spectrum differed from that of 1c by the lack of any proton signal and, moreover, exhibited resonance of the carbomethoxyl protons at 219 Hz, slightly shifted downfield in comparison with the resonance of the same protons in the spectrum

<sup>\*</sup> A check on fresh leaves led to comparable results, though the experimental operations became more difficult owing to appreciable quantities of hydrophilic substances solubilized by water-acetone mixtures

<sup>&</sup>lt;sup>1</sup> Caputo, R, Mangoni, L, Monaco, P and Previtera, L (1974) Phytochemistry 13, 1551

<sup>&</sup>lt;sup>2</sup> CAGLIOTI, L, CAINELLI, G and MINUTILLI, F (1961) Gazz Chim. Ital 91, 1387

<sup>&</sup>lt;sup>3</sup> CHEUNG, H T and FENG, M. C (1968) J Chem Soc (C) 1047

of 1c. Elemental composition and spectral features suggested the structure 2c, in which the double bond is located at the  $\Delta^{13(18)}$  tetrasubstituted position, thus also accounting for the shift of the carbomethoxyl resonance

We have called this substance methyl  $\delta$ -maslinate, in analogy with  $\delta$ -amyrin<sup>4</sup> which has the same tetrasubstituted double bond. The possibility that 2c is an artefact, formed from 1c through acid catalysed isomerization occurring in the isolation procedure, seems to be unlikely. Oleanolic acid (1a) is in fact known<sup>5</sup> not to undergo the acid catalysed isomerization  $\Delta^{12} \rightarrow \Delta^{13(18)}$ , owing to the much faster reversible formation of the corresponding hydroxy lactone. In addition, as far as we are aware, such isomerizations even in the  $\beta$ -amyrin series have been accomplished only in very few cases<sup>6</sup> and then under strongly acidic conditions

The assignment of the structure 2c, based on spectroscopic evidence, was then confirmed by conversion of methyl maslinate (1c) into 2c according to the procedure used by Ruzicka et al.  $^7$  for  $\beta$ -amyrin. Methyl maslinate was in fact acetylated and then refluxed with  $SeO_2$  in acetic acid to afford the  $\Delta^{11,13(18)}$  conjugate diene. Direct hydrogenation of this latter on a prereduced  $PtO_2$  catalyst, followed by alkaline hydrolysis and re-esterification of the carboxyl group, then led to a product m p  $182\text{-}4^\circ$ ,  $[\alpha]_D=26$ , identical with the natural sample

Oleanolic acid (1a), the second substance in the acetone extract, after crystallization from MeOH, has m.p. 307-9,  $[\alpha]_D + 78$  and was identical with authentic material. A check on the mother liquors of crystallization of 1a also showed the enrichment of a second substance, then isolated by further crystallization from hexane. The substance melted over a wide range and was therefore acetylated and esterified, thus being characterized as the corresponding acetoxy methyl ester m.p. 237.9,  $[\alpha]_D - 65$ . The close similarity of its spectral characteristics—mainly the NMR spectrum—with those of 2c immediately suggested structure  $2d^*$ , subsequently confirmed by comparison with authentic 2d obtained from oleanolic acid (1a) as above described for (2b)

<sup>\*</sup> Actually 2d was already described as a synthetic product prepared from methyl oleanolate (1d) 8

<sup>&</sup>lt;sup>4</sup> Musgrave, O. C., Stark, J. and Spring, F. S. (1952) J. Chem. Soc. 4393.

<sup>&</sup>lt;sup>5</sup> Barton, D. H. R. and Holness, N. J. (1952) J. Chem. Soc. 78.

<sup>6</sup> AMES T. R. HALSALL, T. G. and JONES, E. R. H. (1951) J. Chem. Soc. 450.

RUZICKA, L. GROB, A and VAN DER SLUYS-VLIR, F. C. (1939) Helt. Chim. Acta 22, 788

<sup>8</sup> JEGER O. NORYMBERSKE J. and RUZICKA L. (1944) Helt. Chim. Acta 27, 1532

These results show that, in the leaves of *Olea europaea*, maslinic acid exists as such and may be considered a true metabolite; on the other hand, in the fruits of the same plant, it is only formed during the ageing of the husks. Its mode of formation in the husks appears still to be an open question, though the hydroxylation is probably accomplished by a cellular enzyme rather than by a microbial system

#### EXPERIMENTAL

General experimental procedures have been reported elsewhere 1

Extraction of the leaves. Powdered dry leaves (800 g) were continuously extracted with cold hexane for 8 hr and with cold acetone for additional 5 hr. The reddish yellow acetone extract was concentrated, HCl-washed silica-gel (10 g) added and this was evaporated to dryness. The solid residue was then put onto a column filled with HCl-washed silica-gel (530 g). Elution with  $C_6H_6$  (51) afforded a mixture (10 g) consisting of fats and waxes (NMR, GLC), further elution with  $E_2O$  (21) gave a second fraction (6 g) which was treated with 2 N NaOH aq (5 × 100 ml). Acidification of the combined aq layers with conc.  $H_2SO_4$  then afforded a brown waxy mixture (3 g) which, chromatographed on HCl-washed silica-gel, gave three fractions. (A) (0.9 g,  $C_6H_6$ -Et<sub>2</sub>O.9.1) consisting of fats and waxes, (B) (1.5 g,  $C_6H_6$ -Et<sub>2</sub>O.3.2), (C) (0.6 g, Et<sub>2</sub>O)

Oleanolic acid (1a) and acetoxy ester (2d) Fraction (B) was reabsorbed on HCl-washed silica-gel (40 g)  $C_6H_6-Et_2O$  (4 1) eluted oleanolic acid (1a) (1 2 g) m p 307–9° (from MeOH),  $[\alpha]_D+78^\circ$  (c 1 2), identical with authentic material Crystallization from hexane of the mother liquors of 1a gave a solid melting 288–302° which was treated with ethereal  $CH_2N_2$  and then directly acetylated (Ac<sub>2</sub>O-Py overnight) to give the pure acetoxy ester 2d m p 237–9′ (from MeOH),  $[\alpha]_D-65^\circ$  (c 1), (Found C, 77 15, H, 10 17,  $C_{33}H_{52}O_4$  requires C, 77 29, H, 10 22%),  $\delta$  3 65 (-COOMe, s partially overlapping 3-H signal)

Maslinic acid (1b) and methyl  $\delta$ -masliniate (2c) Fraction (C), reabsorbed on HCl-washed silica-gel (20 g) eluting with  $C_6H_6$ -Et<sub>2</sub>O (7 3), gave maslinic acid (1b) (320 mg), mp 267-70° (from MeOH),  $[\alpha]_D + 40^\circ$  (c 0 9), identical with an authentic sample. The mother liquors of crystallization of 1b, treated with ethereal  $CH_2N_2$  and chromatographed on silica-gel (eluent  $C_6H_6$ -Et<sub>2</sub>O 4 1), gave the dihydroxy ester (2c) mp 183-7° (from hexane),  $[\alpha]_D - 28^\circ$  (c 2), (Found  $C_7 = 76.37$ , H, 10.41  $C_{31}H_{50}O_4$  requires  $C_7 = 76.50$ , H, 10.36%),  $v_{max} = 3350$ , 3480 cm<sup>-1</sup>,  $\delta = 365$  (-COOMe, s, partially overlapping 2-H and 3-H signals)

Partial synthesis of 2c Methyl mashinate (1c) (300 mg) was treated with  $Ac_2O$  in anhydrous pyridine overnight and the product was directly dissolved in HOAc (10 ml),  $SeO_2$  (60 mg) added and refluxed for 3 days. Work up of the reaction mixture gave an oil ( $\lambda_{max}^{EioH}$  250 nm)<sup>9</sup> directly submitted to catalytic hydrogenation (PtO<sub>2</sub> catalyst in MeCOOH) for 2 hr. Alkaline hydrolysis (KOH in MeOH 10%) of the product, followed by treatment with ethereal  $CH_2N_2$ , afforded a brown oil which was chromatographed on silica-gel. Elution with  $C_6H_6$ –Et<sub>2</sub>O (4.1) gave a solid (110 mg) m.p. 182-4° (from hexane),  $[\alpha]_D$ –26° identical with natural 2c

Partial synthesis of 2d Methyl oleanolate (1d) was acetylated and the product submitted to the same sequence of reactions described for the preparation of 2c Purification of the final product, by crystallization from MeOH, gave the pure 2d m p  $237-9^{\circ}$ ,  $[\alpha]_D - 65^{\circ}$ , identical with the natural compound

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<sup>9</sup> RAMART-LUCAS, P and HOCH, M J (1935) Bull Soc Chim 2, 327